THE OFFICIAL BIONONYMOUS GUIDE TO

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XTX

MATERIALS



*TIP: You need to measure quantities of 30uL, 100uL, 1mL & 10mL many inexpensive disposable measuring options are sold here: http://www.affwebshop.com/funnels-16-5mm-to-55mm-beakers-pipettes/ **DIY your own centrifuge! http://www.thingiverse.com/thing:1483

STEPS





Put 10mL saline solution per person into individual paper cups. Put 100uL per person of chelex solution into microcentrifuge tubes.







Swirl the cup gently to mix cells that may have settled to the bottom. Use a micropipet with a fresh tip to transfer 1000 µL
of the solution into your labeled 1.5-mL microcentrifuge tube.





Place the sample tubes in a balanced configuration in a microcentrifuge, and spin for 90 seconds at full speed.



Pipet out the supernatant (clear stuff at the top of the tube). Try to remove most of it, but be careful not to disturb the cell pellet (clump of white cells) at the bottom of the tube.





Set a micropipet to 50 $\mu L.$ Resuspend cells in the remaining saline by pipetting in and out.





Withdraw 50 μL of cell suspension, and add it to a tube containing 100 μL of Chelex®. Label the cap and side of the tube.





12.

Place tubes in a balanced configuration in microcentrifuge and spin for 90 seconds at full speed.



Use a micropipet with a fresh tip to transfer 50 μL of the clear supernatant into a clean 1.5-mL tube. Be careful to avoid pipetting any cell debris and Chelex® beads. Label the cap and side of the tube.



Store on ice or in the freezer until ready to use.



15

If you are in a fancy lab quantify your DNA using a nanodrop, Qubit or PCR + gel electrophoresis. If not, try using a capacitance meter:

https://groups.google.com/forum/#!topic/diybio/DCzB2L5iaZo



DNA extraction steps lovingly appropriated from DNALC protocols: www.dnalc.org Written by Heather Dewey-Hagborg, Illustrated by Jarad Solomon